The Immune System as a Reactive System: Modeling T Cell Activation With Statecharts

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Abstract

The construction of reliable reactive systems is considered to be one of the most challenging goals in the fields of software and system engineering. The definition of a reactive system suits biological systems at different levels, ranging from gene networks, developing embryos and the immune system. We report here the application of a tool developed for constructing computerized systems to the modeling and analysis of a biological system, the immune system. We use the language of statecharts within the framework of object-oriented modeling. The results described here indicate that this modeling strategy can contribute to the transition of biology from the phase of analysis to the phase of synthesis.

1. Introduction

There is broad agreement among researchers that biological research must prepare for the transition from *analysis* (the reduction of observations to elemental building blocks) to *synthesis* (the integration of the parts into a whole) [1], and that this transition requires the language of mathematics [2-5]. This is particularly needed in the field of immunology. Over the last several decades there has been an explosion of experimental data describing the cellular and molecular components that comprise the immune system. "At the present time, however, there is an emerging need to understand the system as it functions as a whole" [6].

An illustrative example for the current situation in immunological research is the case of *cytokine networks*. Cytokines are small protein or glycoprotein messenger molecules that convey information from one cell to another. Various aspects of the immune response are regulated by cytokine networks. More than 200 cytokines have now been identified, and more than 12,000 papers on cytokines have been published in 1999 alone. Yet, although many details of particular cytokine interactions have been elucidated, "practically nothing is known about the behavior of the network as a whole" [7].

In this paper we model a biological system using the visual formalism of statecharts [8], as it has been adapted to the framework of *object-oriented modeling* [9-11]. We modeled aspects of T-cell activation. With this model, we are able to run simulations, using the Rhapsody tool [12]. These simulations allow us to compare the dynamic behavior of the model to actual experimental data. Our results show that the use of statecharts can help confront open questions in immunology and probably in other fields of biology, which, because of their complexity, cannot be addressed by standard laboratory techniques alone. In addition, a visual formalism provides a clarity lacking in other approaches to the mathematical modeling of biological systems.

2. The biological system considered

A living cell can be viewed as a system defined by combinations of inner states, reflecting the interactions of the cell with its environment. Transitions between these inner states occur as a consequence of signals that are introduced to the cell by the environment (some of these signals might be produced by the cell itself). Naive T cells, for example, can live for many years without dividing. These naive cells circulate continuously from the blood stream to the lymphoid organs and back to the blood, serving as patrol units that sense changes that might occur in the body. This sensing process is carried out by interactions that the T cells make with antigen-presenting cells. Naive T cells that recognize their specific antigen on the surface of a professional antigen presenting cell cease to migrate, and are activated to proliferate and to differentiate into armed effector cells.

The signaling process that drives cell proliferation can be divided into two stages:

1) "The initial encounter with specific antigen in the presence of the required co-stimulatory signal triggers the entry of the T cell into the G1 phase of the cell cycle and, at the same time, induces the synthesis of IL-2 along with the α chain of the IL-2 receptor" [13].

2) "Binding of IL-2 to the high-affinity receptor then triggers progression through the rest of the cell cycle" [13].

This short description of T cell activation illustrates some of the challenges one encounters while trying to model such a behavior. Describing this behavior raises several questions: Where is the cell? What other entities does it encounter? Which kinds of signals does it receive? What kind of behavioral outputs does it produce? Therefore, we would like to have a modeling language that is able to capture all these various aspects of behavior simultaneously. The need for this parallelism is due to the fact that these various components of behavior are in a sense orthogonal to each other: T cells at the lymph node can be either in an active or in a naive state, and an active T cell can be in any one of the four cell cycle stages.

Another important issue is the ability to distinguish between signals generated outside the cell and signals or events that are produced by the T cell itself. In the above example, the first stage towards cell proliferation – the transition from the G0 stage into G1 – is driven by a combination of two signals that originate from an extracellular event: the interaction of the T cell with a professional antigen-presenting cell. On the other hand, the progression through the rest of the cell cycle is a consequence of two intracellular events: the production of the α unit of the IL-2 receptor and the secretion of IL-2 itself by the T cell.

This traditional description treats the cell a black box, in the sense that it specifies only the outputs produced by the cell in response to given inputs. Nothing is said about *how* the input signals are translated into the outputs; the signal transduction process is totally ignored. Yet, in order to gain a better understanding of such lower level behavior, and to investigate the influence it has on higher level behavior, one would like to have the ability to "zoom" into the system, and describe its inner behavior in a more careful and detailed manner. An adequate modeling approach should support such abilities in a smooth and intuitive, yet formal and rigorous fashion.

3. The Rhapsody tool

Just as programming languages require compilers and debugging tools, so do modeling languages require not only pretty graphics, but also means for executing models and for synthesizing code [14]. In this work, we use the Rhapsody tool (developed by I-Logix), defined as an "object oriented fully integrated Visual Programming Environment (VPE)" (www.ilogix.com), as the working environment. In addition to providing a computerized visual design environment for performing object-oriented modeling, Rhapsody is capable of automatically translating any syntactically legal model into an executable code, currently, in C, C++ or Java. Once the model is constructed and translated into the executable code, Rhapsody can animate the execution of the code so that one can observe its progress in animated versions of the model's statecharts. Animation shows the current state and transitions by coloring them uniquely. In Rhapsody, the transitions between graphical diagrams and executable code are bi-directional: during the compilation stage, the object model diagrams and the statecharts are translated into C++ code lines, and while animating the application, the execution of the C++ code is translated into color changes within the statechart diagrams.

4. Model Construction

4.1 Object Model Diagrams (OMD's)

The first object model diagram (see Fig. 1) describes the relationships between 3 major object classes: *ThelperCell*, *Receptor* and *Ligand*. The *ThelperCell* is a composite object class that contains other kinds of object classes, all of which are *Receptors*. Through these *Receptors*, the T cell can communicate with other objects that belong to the *Ligand* super-class. This is denoted by the 'Bind' relationship, which symmetrically associates *Receptor* and *Ligand*. Another relationship by which the *ThelperCell* class is associated with other objects is the 'Secrete' dependency, which connects the *ThelperCell* to the *Cytokines* class.

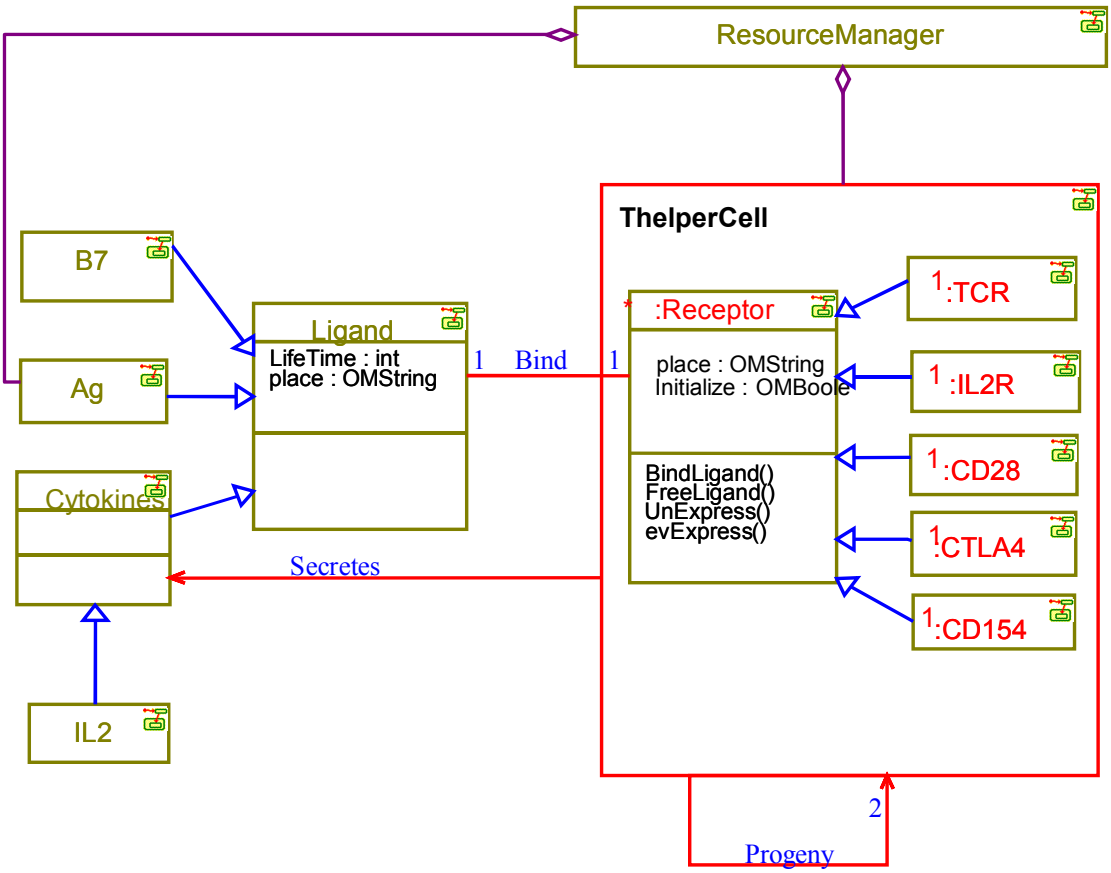

Figure 1

Another dependency arrow named 'Progeny' is drawn in Figure 1 from the *ThelperCell* class to itself, having a one-to-two multiplicity. This relationship implies that a single instance of a *ThelperCell* can refer to two other instances of *ThelperCells* as *its* progeny. This relationship is used during T cell proliferation, when one cell splits into two. We refer to this process as if the mother cell 'dies', and is replaced by two newly born daughter cells.

Another special object class that appears in the object model diagram of Fig. 1 is the *ResourceManager*. The *Resource Manager* carries out three basic functions, essentially 'taking over some tasks from nature', or fulfilling laws of nature:

1. Initiating the first 'generation' of cells (which then can proliferate and die by themselves).
2. Supplying the system with antigen molecules (see also [15]).
3. Manipulating the interactions between receptors and ligands.

Whenever a *Receptor* or a *Ligand* enters a **Free** state, it notifies the *ResourceManager*. The *ResourceManager* acts as a 'matchmaker', continuously searching for a suitable binding partner within the system. If such a match is found, the *ResourceManager* associates the *Receptor* with the corresponding *Ligand*, causing both of them to enter the **Bound** state. In this case, the resource manager carries out *Ligand* diffusion and *Receptor-Ligand* recognition.

It can be seen that all *Receptors* have an instance multiplicity of one in our model. In reality, however, each cell expresses multiple copies of the same receptor. We assume here that when a *Receptor* enters the **Bound** state, it reflects a situation in which the number of receptors bound to a ligand is sufficient to produce an observable response in optimal conditions. Since, on the average, each receptor copy contributes equally to global behavior, we assum that for a qualitative description of the signaling process it suffices to consider the behavior of a single receptor [16].

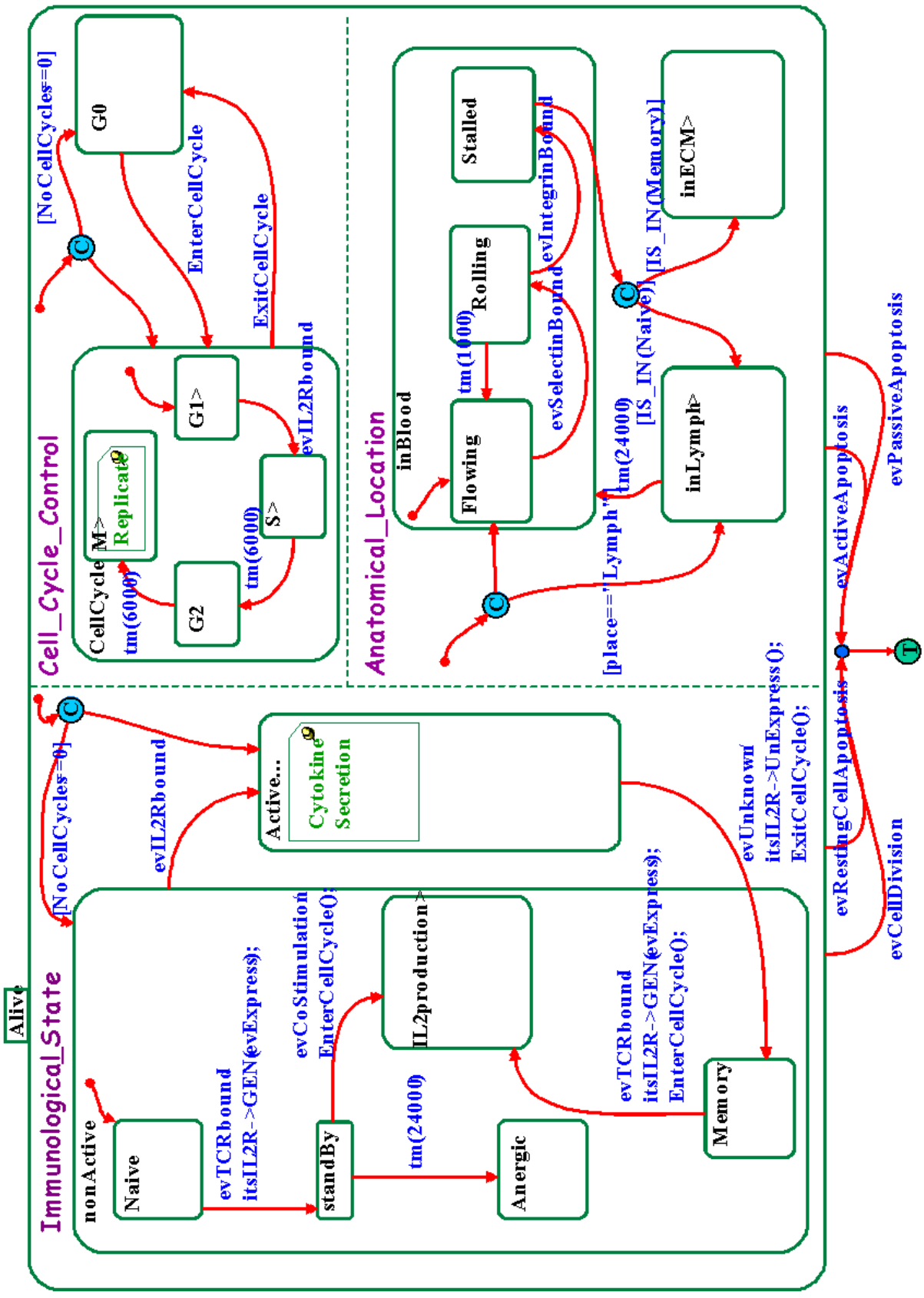
Alive
Immunological_State
nonActive
Naive
evTCRbound
itsIL2R->GEN(evExpress);
standBy
evCoStimulation
EnterCellCycle();
IL2production
tm(24000)
Anergic
evTCRbound
itsIL2R->GEN(evExpress);
EnterCellCycle();
Memory
evUnknown
itsIL2R->UnExpress();
ExitCellCycle();
[NoCellCycles=0]
evIL2Rbound
Active...
Cytokine
Secretion
Cell_Cycle_Control
[NoCellCycles=0]
G0
EnterCellCycle
ExitCellCycle
CellCycle
M>
Replicate
tm(6000)
G1>
S>
evIL2Rbound
G2
tm(6000)
Anatomical_Location
inBlood
Flowing
tm(1000)
Rolling
Stalled
evSelectinBound
evIntegrinBound
[place=="Lymph"]
tm(24000)
[IS_IN(Naive)]
[IS_IN(Memory)]
inLymph>
inECM>
evRestingCellApoptosis
evActiveApoptosis
evPassiveApoptosis
evCellDivision

Figure 2

A second OMD (not shown) extends the system from one cell type to two major cell classes: the T helper cell and the Antigen Presenting Cell (*APC*). The super-class *APC* and the *ThelperCell* are associated by the '**Interaction'** relationship. The conditions under which such interactions can take place are defined later in the framework of the dynamic behavior.

A third OMD (not shown) is dedicated to the *ResourceManager* and relates to all the object classes in the system (since the *ResourceManager* has to be able to 'speak' to all *Receptors* and *Ligands* in order to handle the interactions between them).

4.2 Statecharts: describing dynamic behavior

Since there is a distinct statechart attached to each object class, the current model contains over 20 statecharts. In the interest of clarity, we include detailed descriptions of representative statecharts only, which constitute the core of the model.

4.2.1. The Ligand super-class. Omitted in this version of the paper due to space limitations.

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4.2.2 The ThelperCell class. The statechart of the *ThelperCell* class describes three orthogonal components of the behavior of a T cell (see Fig. 2):

1. Its immunological state — whether the T cell is active or not.
2. Its phase in the cell cycle.
3. Its anatomical location.

4.2.2.1 Immunological state. The immunological state component contains two super-states: **Active** and **nonActive**.

The default transition is conditioned: if the number of cell cycles through which the T cell has gone is zero (i.e., the cell is naive), then the cell will enter the **nonActive** super-state, and within it the **Naive** state. If, however, the number of cell cycles is greater than zero, implying that this newly born cell is the progeny of an active T cell, it enters the **Active** state.

This cumbersome way of charting T cell history is due to the fact that the current version of the Rhapsody tool lacks the capability of *replicating* objects. Currently, it is only possible to create 'fresh' new instances, i.e., ones that start from scratch, in that they enter the statechart's default states. Replication, however, implies that when an existing object instance creates a new instance of the same object class, the statechart of the newly created instance should start in the exact state configuration of the original object at the moment of replication. Such a replication mechanism would enable the creation of instances that carry with them the history of their spawning parents. In order to overcome this current deficiency in the language, we took advantage of the fact that when one object creates a new one (either of its own class, or of any other related object class), the original object can control the initial values of the new instance's attributes. In the case of the T-helper cell object class, these attributes can be made to reflect the relevant history of the T cell, and can be transferred to the daughter cell during proliferation. Therefore, instead of using simple default transitions, we use conditioned default transitions that are dependent on the values of these attributes. These attributes are as follows (in Fig. 2 they appear on the arrows emanating from the condition connectors):

- *NoCellCycles* — the number of cell cycles, which equals zero for naive cells and is greater than zero if the instance was generated during T cell proliferation.
- *Place* — designates the current anatomical location of the T cell. For naive cells, the default location is the blood vessels, while for newly born active cells it is lymphoid tissues.
- *CellType* — indicates whether the cell carries the Th0, Th1 or Th2 phenotype.

Within the **nonActive** super-state, the T-cell can be in one of the following states: **Naive**, **standBy**, **Anergic**, **Memory** or **IL2production**. By default, the T cell enters the **Naive** state. A signal transmitted by the T cell antigen-binding receptor (*TCR*) drives the transition from the **Naive** state to a **standBy** state. During this transition, the T cell carries out another operation – synthesizing the α chain of the IL-2 receptor[1], which is needed for the expression of a high affinity IL-2 receptor (this is done by sending the express event to the T cell's *IL2R* component). The T cell will remain at the **standBy** state until one of the following events occurs:

1. a co-stimulatory signal ('evCoStimulation') is transmitted to it (via the *CD28* receptor);
2. the timeout designated by the 'tm' event expires.

In the former case, the T cell enters the **IL2production** state[2], at which it starts secreting IL-2. In the later case, the T cell enters the **Anergic** state (see below).

This set of transitions stresses the importance of timing, as well as the ability of statecharts to capture the following crucial point: there is a certain 'window of opportunity'

[1]For simplicity, we did not take into account the single chains of the receptor, but merely treated the receptor as expressed (at its high affinity form) or unexpressed. This is justified by the functional aspect – only the high affinity form of the receptor is capable of responding to physiological levels of IL-2.

[2]The requirements for expression of the IL-2 receptor are less stringent than those for IL-2 synthesis. For example, T cell receptor ligation alone is frequently sufficient to induce expression of high affinity IL-2 receptors on T cells. This can allow IL-2 made by one cell to act on IL-2 receptors expressed on neighboring cells [13].

(while the T cell is at the **standBy** state), during which the T cell is capable of responding to the co-stimulatory signal. If this signal is transmitted to the cell just before the antigen binding signal (i.e., while the cell is still in the **Naive** state), or too long after it (when the cell is already in the **Anergic** state), the cell will not be able to respond to the signal. Furthermore, if the cell enters the **Anergic** state, it also loses its ability to respond to future co-stimulatory signals.

It can be seen that while performing the transition from the **standBy** state to the **IL2production** state (as a consequence of the co-stimulatory signal), the T cell also sends a message to itself ('EnterCellCycle') ordering it to enter the cell cycle. The mechanism used for this message is a *triggered operation* rather than an event. This implies that the T cell will not enter the **IL2production** state until the message is processed, ensuring that IL-2 production is synchronized with entry into the cell cycle.

Once in the **IL2production** state, the T cell secretes IL-2. In the current model, each entry into the **IL2production** state induces the creation of one new instance of the IL-2 object class. This newly created IL-2 instance, as a member of the ligand super-class, enters the **Free** state, notifying the resource manager about its state. The resource manager ensures the matching of this IL-2 instance with a free IL-2 receptor, assuming there is such a receptor in the proximity of the *Ligand*. Once bound, the IL-2 receptor transmits the relevant signal to its T cell ('evIL2Rbound'). It can be seen that within the T cell's statechart, this event ('evIL2Rbound') triggers two distinct transitions: the first is from the **nonActive** to the **Active** state within the immunological state component, and the second is from the **G1** to the **S** phase within the cell cycle component.

The **Active** state can be further expanded by the 'open sub-statechart' command, resulting in Fig. 3. The process of T cell activation confronts two basic challenges:

1. *Proliferation*—making more of the same;
2. *Differentiation*—assuming a function: "Late in the proliferative phase of the response, after 4-5 days of rapid growth, these T cells differentiate into armed effector T cells that are able to synthesize all the proteins required for their specialized functions as helper, inflammatory, or cytotoxic T cells" [13]. Subsets of CD4 T cells can be distinguished by their ability to produce discrete patterns of cytokines. The development of these distinct T helper (Th) subsets is strongly dictated by the type of antigen/microorganism invading a host. *Th1 cells*, by their production of IFNγ and lymphotoxin, are responsible for directing cell-mediated immune responses leading to the eradication of intracellular pathogens [17]. *Th2 cells* are excellent helpers for B lymphocytes, and stimulate the production of neutralizing antibodies. The signature cytokines of Th2 cells are IL-4 and IL-5. It is noteworthy that several cytokines produced by Th2 cells (IL-4, IL-10, IL-13) have anti-inflammatory actions. This raises the possibility that an important physiological function of Th2 cells is to serve as regulators of immune responses that limit the injurious consequences of Th1-mediated protective immunity [18]. *Th0 cells* express some differentiated effector functions of both the inflammatory and anti-inflamatory T cells. Whether Th0 cells are precursors for Th1 and Th2 cells or represent a separate, stably differentiated population remains unclear [17].

Control of proliferation is carried out within the cell cycle component of the statechart. In the current model, we do not get too deeply into the differentiation aspect of T cell activation. However, a basic sketch appears in the **Active** sub-statechart (see Fig. 3).

As helper T cells differentiate, they are thought to go through an intermediate stage known as Th0. Therefore, the default arrow in the **Active** sub-statechart leads to the **Th0** state. Further differentiation into Th1 or Th2 armed effector T cells is dependent on two basic factors:

1. The cytokine pattern sensed by the T cells. Two key cytokines that control this process are IL-12 and IL-4, which drive the cells towards the Th1 and Th2 pathways, respectively.
2. The number of cell cycles these cells have gone through. Th0 cells fail to respond to IL-4 unless they have traversed at least 3 cell cycles, while the responsiveness of Th0 cells to IL-12 was found to increase as cell proliferation proceeds: "A naive cell can become a Th1 cell with increasing likelihood after each successive cell cycle but can only become a Th2 cell when eight siblings have arisen" [19]. This dependency on cell cycle progression was found to be related to cell cycle dependent changes in chromatin

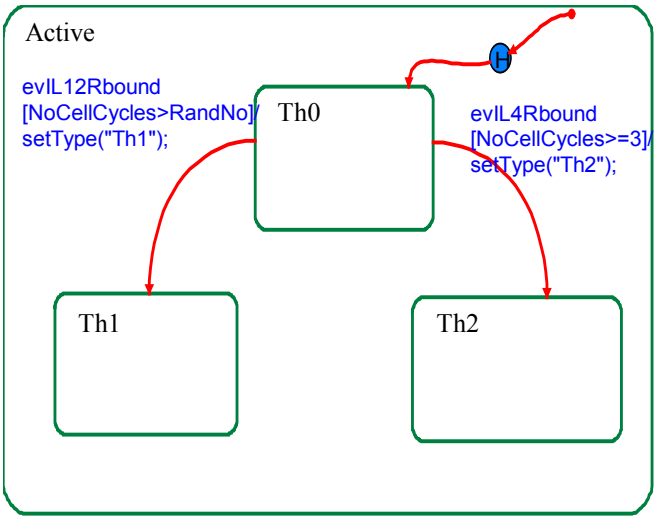

Figure 3

structure, which gradually 'expose' these cytokine genes to transcription.

In the model, the combination of these two factors is taken care of by the guarded transitions emanating from the **Th0** state (see Fig. 3). The **Th0**-to-**Th2** transition will take place when the event of binding IL-4 is transmitted by the IL-4 receptor ('evIL4bound'), only if the condition *'NoCellCycles'* ≥ *3* is fulfilled. As for the **Th0**-to-**Th1** transition, the conditional guard here adds a probability factor to the transition: A random number between 1 and 5 is chosen; if this number is smaller than the number of cell cycles, the transition will take place once the event of IL-12 binding is generated by the IL-12 receptor. This mechanism ensures that the probability of the IL-12-binding event to promote the Th1 state will increase with the number of cell cycles.

There are three different circumstances under which the T cell might enter the **Active** state:

1. After the initial encounter with an antigen, followed by either the appropriate co-stimulatory signal or by the binding of exogenous IL-2.
2. If the T cell is 'born' during proliferation, the **Active** state is entered by default.
3. If the T cell is a memory cell, which, as a consequence of re-encountering its antigen, becomes re-activated.

In the first case, the default transition into **Th0** takes place, since the system has no history of activation. In the third case, the cell enters the same state in which it resided the last time it left the **Active** state: For example, a memory T cell that entered the **Th1** state when it first became an effector cell would once again enter the **Th1** state. The same holds for Th2 and Th0 T cells. Concerning the second case, an active T cell generated during proliferation should adopt at 'birth' the same conformation as its mother cell. However, as mentioned earlier, there is no current mechanism in Rhapsody that supports object replication. Hence, we check the value of attributes that the mother cell transfers to the T cell in question. This is done using the *Type* attribute, which can have one of three string values 'Th0', 'Th1' or 'Th2'[3].

As for the immunological state componenet, there are two additional states in which a *ThelperCell* can be found (Fig. 2):

Anergic state[4]: "Naive T cells that recognize antigen on cells that cannot provide co-stimulation become anergic, as when a T cell recognizes a self antigen expressed by an uninfected epithelial cell. This T cell does not differentiate into an armed effector cell, and cannot be stimulated further by professional antigen-presenting cells" [13]. This possibility is represented by the transition from the **standBy** state to the **Anergic** state, taking place if no co-stimulation event is provided during a certain time delay after the antigen-binding event. It can also be seen that the only possibility of 'escaping' the **Anergic** state is if the IL-2 receptor gets bound, resulting in a transition to the **Active** state. There are reports about IL-2 rescue of anergic T-cells [20], which could be explained by results that show that TCR signaling is sufficient for the expression of the IL-2 receptor[13].

1. **Memory** state: "Immunological memory can be defined as the faster and stronger response of an animal that follows re-exposure to the same antigen" [21]. The events that trigger the differentiation of an active effector T cell into a memory cell are still unknown. Yet, "irrespective of how memory CD4 T cells develop, they are the progeny of naive T cells that have successfully recognized foreign peptide in association with MHC class II… the generation of memory may be envisaged as the maturation of antigen-primed naive CD4 T cells" [22]. Memory T cells are long-lived lymphocytes that remain in a resting state after the antigen has been cleared. During the transition from the **Active** state to the **Memory** state, there is the complete loss of the surface adhesion molecule L-selectin, which mediates the homing of T cells to the lymph nodes. However, "unlike naive CD4 T cells, memory cells can enter non-lymphoid tissues directly from the blood" [22], thus enhancing the chance of antigen encounter (see also the anatomical location component in Fig 2). In order to become re-activated, the resting memory T cell must re-encounter its antigen, presented by an APC.

As for further requirements, things are not so clear: "The role of co-stimulation in regulating memory CD4 T cell responses is still highly controversial" [22]. It has been shown that, unlike naive T cells, resting memory cells respond to antigens presented by APC's in the absence of co-stimulation. Other studies, in apparent contradiction, suggest that a secondary response might be blocked by preventing co-stimulatory signals. In our model (Fig. 2), we include the first possibility, as can be seen by the fact that the 'evTCRbound' event triggers the immediate transition from the **Memory** state to the **IL2production** state. However, Fig. 4 shows an alternative description, in which the **Naive** and **Memory** states are combined into a **Resting** state, from which the 'evTCRbound' event leads to the **standBy** state; only if a further co-stimulatory event occurs during the defined time delay, will the activation pathway take place.

[3] Actually, once we have these conditional transitions, we do not really need the history connector. Still, we wanted to illustrate the usefulness of such historical memory, assuming that the problem of object replication can be solved.

[4] We didn't take into account anergy induced by binding of altered peptide ligands (APLs, antigens that bind to the TCR with low affinity). This is due to the fact that we have not dealt with binding affinities.

4.2.2.2 Cell cycle control. As mentioned above, naive T cells can live for many years without replicating, implying that during this time period they stay in the **G0** state. The initial encounter with a specific antigen in the presence of the required co-stimulatory signal triggers the transition from **G0** to the **G1** phase of the cell cycle. In our model, the transition from the **G0** state to the **G1** state is driven by the 'EnterCellCycle' triggered operation, which is initiated during the transition from the **standBy** state to the **IL2production** state. Further progression through the cell cycle is dependent on IL-2 binding by the IL-2 receptor, an event that triggers both the transtion from the **G1** state to the **S** state in the *Cell_Cycle_Control* component and the transition from the **nonActive** state to the **Active** state in the *Immunological_State* component. If no death signal is induced while being in the **S** phase of the cell cycle, the cell will go through the rest of the cell cycle, depending only on timeout events. Once entering the **M** state, a series of actions is carried out, resulting in the generation of two new instances of the ThelperCell class, representing the two newly born daughter cells.

4.2.2.3 Anatomical location. Here, we have not provided a detailed description of T cell migration. The main role of the anatomical location component is to provide the spatial context for receptor-ligand interactions: in order for a chemical interaction to take place, the two interacting entities must be in proximity. Whenever a T cell changes its location, it informs the resource manager about this transition. The *ResourceManager* then searches among the relevant free *Ligand* objects for such that share with the T cell the same anatomical location.

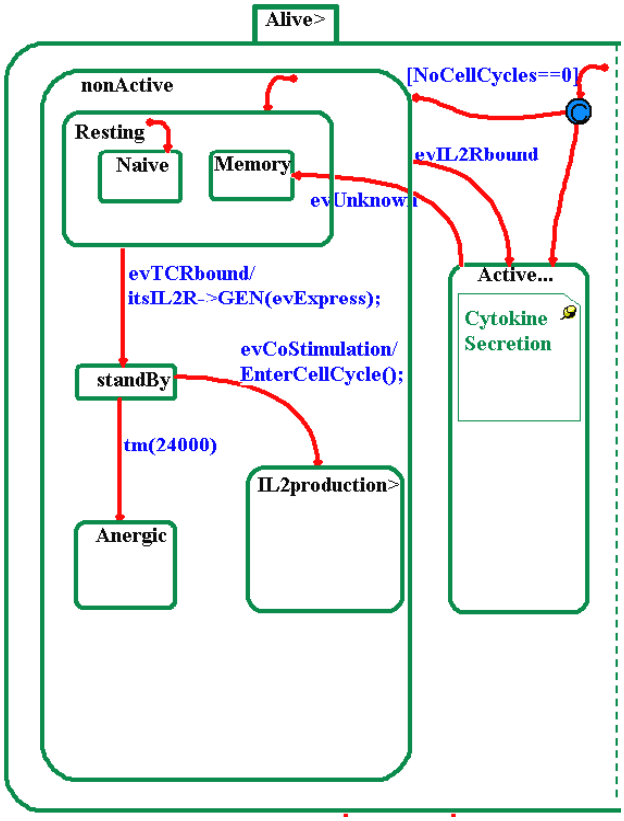

Figure 4

5. Results

5.1 Model execution

In order to check whether the formal representation of the model fulfills our requirements (based on the immunological data presented in previous sections), we executed the model using the Rhapsody tool. Due to space limitations, we omitted this section from this version of the paper; this section will be included in a more comprehensive version, which is in preparation.

5.2 Unexpected Behavior

By and large, the behavior we encountered in the executions of the model followed our expectations, except for one case: we found that the *Thelpercell* object could not reach a steady **Memory** state. The moment a T cell took the transition from **Active** to **Memory**, it returned right back to the **Active** state. This happened because IL-2 molecules were still in the system, while the *Thelpercells*, being in their **Active** state, expressed their *IL2R* component objects. As shown in Fig. 2, the event of IL-2 binding by the IL-2 receptor triggers the transition from the **nonActive** super-state to the **Active** state. Only after an extensive search in the literature did we find that the transition from **Active** to **Memory** is accompanied by downregulation of IL-2 receptor expression, thus causing the memory T cells to become insensitive to the presence of IL-2 [23]. Accordingly, we corrected the statechart by adding the action 'itsIL2R->GEN(evUnExpress)' to the transition leading from the **Active** state to the **Memory** state (see Fig. 2).

6. Discussion

In this work, we examined the feasibility of statecharts in bridging the gap between analysis and synthesis by modeling a well-studied immunological process termed T cell activation using the language of statecharts, as implemented in the framework of the Object-Oriented Analysis and Design (OOAD) approach. This methodology supports the following attributes:

Concurrency: Biological systems in general, and the immune system in particular, are highly concurrent. Changes in the surrounding environment can trigger the

execution of many parallel processes. At the cellular level, a given cell carries out different tasks in parallel, as shown in Fig. 4 (e.g., secreting IL-2 and progressing though the cell cycle). Dividing the T-cell's statechart into three orthogonal components enabled us to derive a description of T-cell behavior which is composed of three different aspects: the T-cell's immunological state, its cell-cycle phase and its anatomical location.

Multi-level description: Information processing in biological systems occurs at different levels, ranging from the molecular and cellular levels, up to the higher levels of organisms and populations. Computations in biological systems are carried out on each of these levels. At the molecular level, for example, the T-cell receptor could be simply regarded as **Free** or **Bound**. One can further zoom into the sub-molecular level, and divide the binding process into several events occurring at the sub-molecular level: successive additions of phosphate groups to six different positions on the zeta (ζ) chain of the antigen receptor (TCR) [24]. Only a full phosphorylation of all six sites will result in an activation signal transmitted by the TCR. Completion of the phosphorylation steps is dependent on the nature of the TCR ligand. Thus, the phosphorylation steps establish a threshold for T cell activation at the sub-molecular level.

As well as traveling between the molecular and sub-molecular description level, a multi-scale description is also needed at the cellular and sub-cellular levels. The model described in this paper, for example, treats the T cell as a 'black box': it describes the output behavior of the cell as a reaction to external signals, regardless of the signal transduction events that occur within the cell and lead to this behavior. Thus, the model simply states that proper signaling via the TCR, followed by co-stimulation, triggers the transition from **Naive** to **Active**. The present model says nothing about the molecular or sub-molecular meaning of "proper signaling" (the phosphorylation events), neither does it take care of the cascade of events that translate the signal transmitted by the bound TCR into a transition from **Naive** to **Active** (the signal transduction pathway). The model, however, can be modified to include signal transduction events as well (though we have not done so in this paper). This 'zooming in' process has two implications: (i) adding new components (e.g., *enzymes*) to the *ThelperCell* composite object, and (ii) adding several orthogonal components to the *ThelperCell's* statechart – a distinct component for each signal transduction pathway.

This ability to describe the system in a multi-scale manner answers one of the key challenges of theoretical biology: "understanding how detail at one scale makes its signature felt at other scales, and how to relate phenomena across scales" [4].

The model described in this work was constructed in the framework of the **object-oriented** modeling approach. This methodology was favored for the following reasons:

- Describing the system as a collection of objects that communicate between them highly correlates with the challenge of studying entire biological systems "by attempting to understand how component parts collaborate to create a whole" [3].
- Interestingly, most of the experimental data in biology accumulates in an object-oriented manner. The specialization through which biological research has progressed during the last few decades has lead to the current situation whereby most research has been reduced to questions about "the individual components of complex biological systems". As a consequence, much is known about individual proteins (their amino acid sequence, their 3D structure, the role they play in concrete biological reactions, the identity of other proteins with which they interact, etc.), but much less is known about networks of genes and proteins. Concerning the immune system, a significant amount of data exists about its cellular compartments (the receptors they express, the molecules they secrete, their activation requirements, etc.), but very little is known concerning the way these cells collaborate to function as a system. Object-oriented modeling provides a useful formalism for constructing models by organizing information gathered around individual objects.
- People actually think about biological systems in an object-oriented manner. The first section of the immunology textbook of Janeway & Travers is entitled "The components of the immune system" [13]. This reflects our tendency to base most of the discussions related to biological systems and processes on the concrete biological objects that constitute the system.

These three issues are related to each other. Yet, presenting them as three distinct points illustrates the great advantage of using the object-oriented modeling approach: it fits the way we think, it fits the way the experimental data are collected and it seems suitable for coping with the challenge of understanding how biological objects collaborate to establish a system.

As shown in this work, statecharts can be used to represent different aspects of biological system behavior, at varying scales of description. Constructing statecharts-based models can be helpful in addressing various questions, such as:

- How can the order of events and the duration of time delays influence the behavior of the system?
- Can contradictory output behaviors be explained by reorganizing inner states into different clusters or orthogonal components? This was shown here for the **Memory** state, where two alternative clustering

possibilities were suggested for representing contradictory observations related to costimulatory requirements (Figs. 2, 4).

Besides predications and explanations, "one of the beauties of mathematical modeling is that it raises questions that may not have been addressed before" [6]. Merely organizing the existing information using a visual formalism can raise questions that were not thought of before.

Finally, modeling can influence not only the biological system, but also the computational methods used. Confronting tools developed for modeling and analyzing industrial systems with the challenge of modeling and analyzing biological systems may reveal limitations of the computational tools that were not appreciated before. During our modeling work, two such points became evident:

1. Currently, the Rhapsody tool lacks a mechanism for object replication. As a consequence, modeling the process of T cell proliferation resulted in a relatively cumbersome model (see Section 4.2.2.1).
2. Communication between objects in Rhapsody is handled as point-to-point communication, i.e., each message sent by a given object instance has only one destination (the address of the recipient object instance). Rhapsody does not allow *broadcasting*, i.e., sending messages to groups of objects, such as all the components of a composite object (limited broadcast) or all the objects in the system (full broadcast). The lack of a broadcast mechanism in Rhapsody is the main reason for heavily relaying, in the present model, on the *ResourceManager* for enabling *Ligand-Receptor* interactions (see Section 4.1).

These and other deficiencies of the modeling language/tools can be rectified. But even at this early stage of application, it is rewarding to see how well an object-oriented visual language fits the needs of biological understanding.

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